

PI Name:

Safety Protocol for Safe Handling and Disposal of Biosafety Level 1 Microbes

1. Hazard Communication Statement

Strains of BSL1 yeast such as *Saccharomyces cerevisiae* and *Schizosaccharomyces pombe* are used in the brewing industry convert the sugar in grain or fruit to alcohol. Brewer's yeast dies when alcohol concentrations exceed its tolerance level of about 16 percent. Baker's yeast, a live strain of the same fungus, causes intestinal gas and thiamine deficiency if eaten as the raw powder. Neither baker's nor brewer's yeast causes vaginal or skin infections. There is a very limited record of adverse effects to human health or the environment. Lab strains of *S. cerevisiae* and *S. pombe* are classified as biosafety 1 level organisms and are frequently used as model organisms in cell and molecular biology. The yeast species *S. pombe* and *S. cerevisiae* are both well studied; these two species diverged approximately 600 to 300 million years ago, and are significant tools in the study of DNA damage and repair mechanisms. Based largely on the extensive history of safe use, many experiments with yeast are exempt based on NIH guidelines. *S. cerevisiae* may result in colonization of the body only with the ingestions of high levels of the microbe or following exposure of immune suppressed individuals. In rare circumstances, *S. cerevisiae* can cause fungemia in hospitalized patients.

Nickoloff, Jac A.; Haber, James E. (2011). "Mating-Type Control of DNA Repair and Recombination in *Saccharomyces cerevisiae*". In Nickoloff, Jac A.; Hoekstra, Merl F. *DNA Damage and Repair*. Contemporary Cancer Research. pp. 107–24. ISBN 978-1-59259-095-7.

Popiel KY, Wong P, Lee MJ, Langelier M, Sheppard DC, Vinh DC. [Invasive *Saccharomyces cerevisiae* in a liver transplant patient: case report and review of infection in transplant recipients](#). *Transpl Infect Dis*. 2015 Jun;17(3):435-41. PMID: 25827213

Muñoz P, Bouza E, Cuenca-Estrella M, Eiros JM, Pérez MJ, Sánchez-Somolinos M, Rincón C, Hortal J, Peláez T. [Saccharomyces cerevisiae fungemia: an emerging infectious disease](#). *Clin Infect Dis*. 2005 Jun 1;40(11):1625-34. PMID: 15889360

2. Laboratory Precautions

2.1. Standard Laboratory practices

Microbes should be handled with appropriate precautions consisting primarily of good microbiological laboratory techniques. The following precautions should be employed:

- A. Access to the laboratory is limited or restricted at the discretion of the laboratory director.
- B. Do not store food in lab.
- C. Do not eat, drink, smoke, handle contact lenses, apply cosmetics (including chap stick), etc. in the lab.
- D. Do not mouth pipette.
- E. Laboratory personnel must be appropriately trained.
- F. The safety protocol (SOP) serves as training documentation and reference information. A copy signed by laboratory personnel should be kept in the lab's safety manual.
- G. Infectious or biohazardous materials must be transported in sealed primary container inside a sealed durable and leak proof secondary containment labeled with a biohazard sticker.
- H. Liquids should be handled carefully to minimize creation of splashes and aerosols. Centrifugation should be performed using sealed tubes and sealed rotors.
- I. Sharps should be handled with extreme caution to avoid cuts or autoinoculation during use and disposal. Needles should not be bent, sheared, or recapped. The needle and syringe should be promptly placed in a puncture-resistant container and decontaminated, by autoclaving or incineration.
- J. Decontamination:
 - i) Liquid: cultures and liquid waste by adding bleach to a final concentration of 10% bleach for a minimum of 30 minutes.
 - ii) Solid waste: Dispose of solid wastes (such as Petri dishes, inoculating loops, disposable centrifuge tubes, pipettes and gloves) in orange biohazard bags, which are autoclaved and placed in red biohazard bags/containers for final disposal.

- iii) Surfaces: Decontaminate work surfaces with 70% ethanol or 10% bleach (made fresh weekly) after a spill and when work is completed for the day.
- iv) Materials to be decontaminated outside the lab must be placed in a durable leak proof container and secured for transport
- K. Lab personnel must wash their hands after they handle viable materials and animals, after removing gloves, and before leaving the laboratory or animal facility.

2.2 Personal Protective Equipment (PPE)

- A. Personnel are encouraged to keep a change of clothes in lab in the event their clothes become contaminated.
- B. Wear lab coats and gloves when working with bacterial cultures.
- C. Wear safety glasses when splashes sprays or aerosols can be expected.
- D. Dispose of contaminated gloves and disposable lab coats in biohazard bags/containers.
- E. Contaminated cloth lab coats may be disinfected by soaking in 10% bleach solution for 30 minutes.
- F. No personal protective equipment is to be worn outside of the lab.

2.3 Bunsen burner precautions (if applicable)

Bunsen burners pose an obvious fire hazard. Proper adjustment and use are important. Please observe the following precautions when using a Bunsen burner.

- A. Locate the fire extinguisher prior to igniting the Bunsen burner.
- B. Clear your lab bench of any clutter and debris that might accidentally catch fire. Keep containers of alcohols and other flammables at a safe distance.
- C. Keep the burner on only when in use and be mindful of the flame, which can burn quietly and may appear almost invisible under fluorescent lights. Individuals may forget the burner is ignited and inadvertently pass an arm or lean over it.
- D. Adjust the burner's gas flow and air intake to ensure optimum performance.
- E. Individuals with long hair may be at increased risk of accidentally singeing or burning their hair and shall reduce this risk by tying back or in some way bidding their hair.
- F. Note: The fire hazard associated with Bunsen burners can be mitigated by utilizing electric microincinerators rather than traditional Bunsen burners.

3. Emergency procedures

3A. Spills of Microbial Cultures:

- 1) Notify workers in the area.
- 2) Leave the area for 15 minutes to allow aerosols to settle. Replace contaminated PPE.
- 3) Upon return, mix spill with freshly made bleach to 10% final concentration.
- 4) Allow 30 minutes of contact time for disinfection.
- 5) Absorb spill with paper towels and dispose them into biohazard bags.
- 6) Use dustpan and broom to sweep up debris. Broken glass must be deposited into broken glass or sharps box.
- 7) Wipe the spill area clean using 10% bleach.
- 8) Dispose of contaminated PPE in autoclavable biohazard bags.

3B. In the event of injury or exposure

- 1) **CLEANSE WOUND:** Wash all wounds immediately with antiseptic soap and a high volume of water for up to 15 minutes.
- 2) **CONTROL BLEEDING**
- 3) **ACCIDENTAL INGESTION:** Rinse mouth with water but do not swallow.
- 4) **SEEK IMMEDIATE MEDICAL FOLLOW-UP (do not wait 24 hrs)**

Employees *and* students go to:

- **Employee Health Services** (during business hours: Monday-Friday, 7:30 am -4 pm).
Address/Location: 57 Bee Street, Charleston SC 29425; Phone: (843) 792-2991
- **MUSC Emergency Room** (after business hours)

